

Thermodynamic and kinetic approaches for drug discovery to target protein misfolding and aggregation

Michele Vendruscolo

To cite this article: Michele Vendruscolo (2023) Thermodynamic and kinetic approaches for drug discovery to target protein misfolding and aggregation, Expert Opinion on Drug Discovery, 18:8, 881-891, DOI: [10.1080/17460441.2023.2221024](https://doi.org/10.1080/17460441.2023.2221024)

To link to this article: <https://doi.org/10.1080/17460441.2023.2221024>



© 2023 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 05 Jun 2023.



Submit your article to this journal [↗](#)



Article views: 1430



View related articles [↗](#)



View Crossmark data [↗](#)

REVIEW



Thermodynamic and kinetic approaches for drug discovery to target protein misfolding and aggregation

Michele Vendruscolo 

Centre for Misfolding Diseases, Yusuf Hamied Department of Chemistry, University of Cambridge, Cambridge, UK

ABSTRACT

Introduction: Protein misfolding diseases, including Alzheimer's and Parkinson's diseases, are characterized by the aberrant aggregation of proteins. These conditions are still largely untreatable, despite having a major impact on our healthcare systems and societies.

Areas covered: We describe drug discovery strategies to target protein misfolding and aggregation. We compare thermodynamic approaches, which are based on the stabilization of the native states of proteins, with kinetic approaches, which are based on the slowing down of the aggregation process. This comparison is carried out in terms of the current knowledge of the process of protein misfolding and aggregation, the mechanisms of disease and the therapeutic targets.

Expert opinion: There is an unmet need for disease-modifying treatments that target protein misfolding and aggregation for the over 50 human disorders known to be associated with this phenomenon. With the approval of the first drugs that can prevent misfolding or inhibit aggregation, future efforts will be focused on the discovery of effective compounds with these mechanisms of action for a wide range of conditions.

ARTICLE HISTORY

Received 23 January 2023
Accepted 30 May 2023

KEYWORDS

Protein misfolding; protein aggregation; protein phase separation; amyloid aggregates

1. Introduction

The phenomenon of protein misfolding and aggregation has been associated with over 50 degenerative diseases, collectively known as amyloidoses [1–3]. These conditions include neurodegenerative disorders such as Alzheimer's and Parkinson's diseases, systemic disorders such as light chain (AL) amyloidosis and hemodialysis-related amyloidosis, and localized disorders such as type II diabetes [1–3]. The proteins that misfold and aggregate in these diseases are unrelated in sequence, structure and function [1,4]. Some proteins, including amyloid β (A β) [5,6] and tau [7,8] in Alzheimer's disease, α -synuclein in Parkinson's disease [9,10], and IAPP in type II diabetes [11,12], are natively disordered. Other proteins, including antibody light chains in amyloid light-chain amyloidosis [2,13], β_2 -microglobulin in hemodialysis-related amyloidosis [14] and transthyretin in familial amyloid polyneuropathy [15,16] are folded in their native states.

Although the clinical manifestations of these diseases cover a vast range of symptoms, they share a common molecular signature, as they are associated with the presence of amyloid aggregates in the affected tissues and organs [1–3,17]. The amyloid state is characterized by the formation of long fibrillar assemblies with a core structure composed of β -sheets whose strands run perpendicular to the fibril axis [1,17], and stabilized by a network of backbone hydrogen bonds [18]. The advent of cryo-electron microscopy has recently enabled the determination of patient-derived high-resolution amyloid structures associated with many different protein misfolding

diseases, revealing the existence of different disease-specific amyloid folds, often referred to as fibril polymorphs [19–22].

In order to develop effective treatments based on amyloid aggregation as a target, one should understand the molecular origins of this process, as well as characterize and quantify the most cytotoxic aggregated species produced during the formation of the amyloid state [1–3,6,23,24]. It has been proposed that supersaturation is the common driving force for the aggregation of these proteins [25,26]. In this view, many proteins are more stable in their amyloid states than in their native states, at least at the concentrations at which they are expressed under normal cellular conditions [25,26] (Figure 1). Under these circumstances, aggregation can be delayed but cannot be completely avoided. Proteins can perform their functions because their native states are kinetically more accessible than the amyloid states, and high free energy barriers slow down the aggregation process [1,27,39]. Furthermore, when they form, protein aggregates are removed by degradation pathways, including the ubiquitin-proteasome and the autophagy-lysosomal systems [40]. More generally, the protein homeostasis system regulates the behavior of proteins in terms of their conformations, interactions, concentrations and localization [41,42], and contributes to the maintenance of the functional state of proteins in the presence of a thermodynamic drive toward aggregation [40]. Upon aging, however, the progressive impairment of the protein homeostasis system may lead to the gradual accumulation of protein deposits, whose presence in turn initiates a cascade of pathological processes [43–49].

Article highlights

- Protein misfolding and aggregation into the amyloid state is a widespread phenomenon, which, when dysregulated, results in a variety of different diseases.
- At physiological protein concentrations, the amyloid state can be thermodynamically more stable than the native state. Under these conditions, aggregation is inevitable, although proteins should cross high-free energy barriers to aggregate.
- Thermodynamic approaches for drug discovery for protein misfolding diseases are based on the stabilization of the native state with respect to the misfolded and amyloid states.
- Kinetic approaches for drug discovery to target protein aggregation are based on reducing the rate of conversion between the native and amyloid states.
- Thermodynamic and kinetic approaches could be extended to amyloid aggregation within liquid-like condensates generated by protein-phase separation.

Therapeutic interventions could thus be aimed at supplementing the protein homeostasis system as it is progressively impaired upon aging. In broad terms, one could think of two complementary strategies to reduce the driving force toward aggregation, the first based on the thermodynamics and the second on the kinetics of the process (Figure 1). In a thermodynamic strategy, since

supersaturation depends on the free energy difference between the native and amyloid states, the aim is to develop compounds that stabilize native states [16,28–32]. In a kinetic strategy, the goal is to increase the height of the free energy barrier between the native and amyloid states. In this second approach, the thermodynamic driving force remains unchanged, but the process is slowed down, so that even a protein homeostasis system of reduced functionality could effectively remove the aggregates [33–38]. We should point out that the optimization of the kinetics of binding of small molecules to their targets, in terms of the balance between on rates and off rates, has a long history in drug discovery [50–52]. However, the type of kinetic strategy that we are concerned with here is specifically aimed at increasing the rate of conversion of native states into aggregates.

In this review, we discuss recent results obtained using the thermodynamic and kinetic strategies against protein aggregation.

2. Thermodynamic approaches

A drug discovery strategy to target protein aggregation based on the stabilization of the native state was first demonstrated

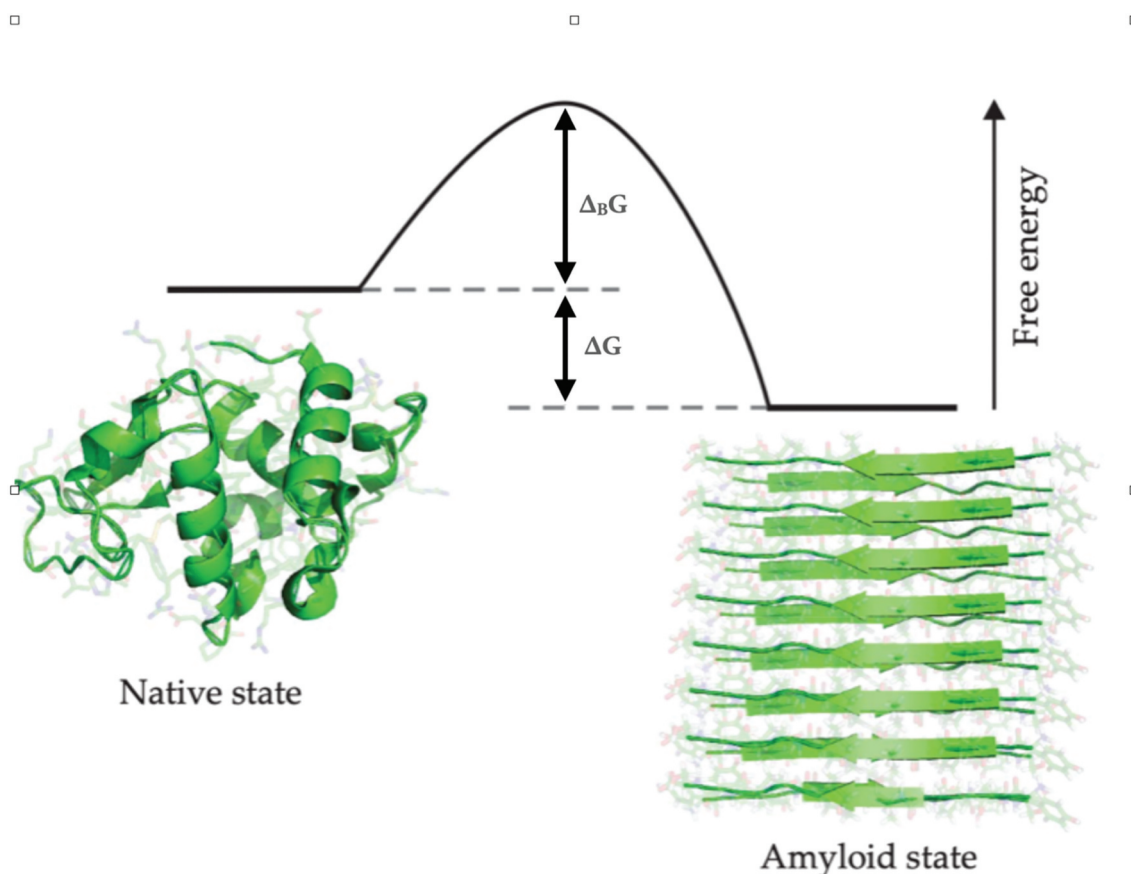


Figure 1. Schematic free energy landscape of protein aggregation. at the concentrations at which proteins are expressed, the amyloid state of proteins can be thermodynamically more stable than the native state [26,27]. Thermodynamic approaches to drug discovery are aimed at stabilizing the native state by reversing the sign of the free energy difference (ΔG) between the native and amyloid states [16,28–32]. Kinetic approaches have instead the target of increasing the free energy barrier for the conversion between the two states (Δ_{BG}), thus prolonging the lifetime of the native state [33–38]. Thermodynamic and kinetic approaches can be complementary to each other. The former may be more effective at preventing aggregation at low protein concentrations, while the latter may be preferable at higher concentrations where the aggregation process is more rapid. Ultimately, the choice of approach depends on the specific circumstances of the protein aggregation problem at hand. Reproduced from [1] with permission of Springer Nature.

over a decade ago in the case of transthyretin, initially for familial amyloid polyneuropathy, and subsequently for both familial and sporadic amyloid cardiomyopathy [16,28,53,54]. Starting from the observation that the first step in the aggregation pathway of transthyretin is the dissociation of the native homotetramer, the small-molecule tafamidis was identified to stabilize the native complex [28,53].

Based on the example of tafamidis, other compounds with a similar mechanism of action, known as pharmacological chaperones [16,55], have been developed. Six drugs for three other amyloidoses have been recently approved by the FDA: (i) migalastat, which targets destabilized mutant α -galactosidase A in Fabry disease [56], (ii) voxelotor, which targets a mutant form of hemoglobin in sickle cell disease by stabilizing the tetrameric oxygenated form of the protein [57], and (iii) ivacaftor (also known as VX-770) [58], elexacaftor (VX-445) [59] and tezacaftor (VX-661) [60], which act synergistically to target mutant forms of the anion channel cystic fibrosis transmembrane conductance regulator (CFTR) in cystic fibrosis by promoting channel gating (ivacaftor) and by stabilizing the native state (elexacaftor and tezacaftor) [16,59]. Other compounds with a similar mechanism of action are under development. A small molecule in the sterol class has been shown to act as pharmacological chaperone by inhibiting amyloid fibril formation of mutant α -crystallins, and to reverse cataract in mouse models of these mutants [61,62]. Another sterol (lanosterol) was shown to ameliorate cataract in dogs [63], and is currently commercialized as a veterinary drug (Lanomax, <https://www.lanomax.com/>). Pharmacological chaperones are also being investigated with great promise to target oncogenic mutants of the tumor suppressor protein p53 [64].

By building on these developments, it has been proposed that approaches based on the stabilization of the native states of proteins may be extended to disordered proteins, which until recently have been considered undruggable [29,31,65–69]. Finding effective drug discovery strategies for this class of proteins would have a major impact since thousands of potential targets are disordered proteins, which are involved in essentially all major human diseases [29,67].

The major obstacle to achieve this goal is that disordered proteins do not readily exhibit binding pockets for forming stable interactions with small molecules. Nevertheless, small molecules have been identified to bind disordered proteins using a range of methods, including nuclear magnetic resonance (NMR) spectroscopy, surface plasmon resonance (SPR), biolayer interferometry (BLI), and fluorescence techniques [66,70–72]. It has been suggested that small molecules may induce the folding of a disordered protein, perhaps just to the extent in which well-formed binding pockets are formed [66,73,74]. It is not yet clear, however, whether drug-like affinity and specificity can be achieved through this type of binding mechanism. To obtain higher affinity and specificity, one possibility is to use larger molecules, such as macrocycles, peptides, or antibodies [75–77]. However, for brain diseases, or diseases in which protein aggregation is intracellular, small molecules would remain preferable, as they would more readily cross the blood-brain barrier and the cell membrane.

Another possibility, beyond the strategies based on the existence of binding pockets mentioned above, is to exploit a different binding mechanism, one in which the complex between the

protein and the small molecules remains disordered. This mechanism of binding has been observed for protein–protein interactions and protein–nucleic acid interactions [78–80], and it is likely to be responsible for the phenomenon of protein phase separation (PPS), which has been associated with the formation of membraneless organelles [81–85].

The possibility of a disordered binding mechanism for small molecules has been illustrated recently in the case of A β , where the small-molecule 10074-G5 was shown to bind and stabilize the disordered native state of this peptide, thus preventing its aggregation [29,31,32] (Figure 2). The nature of the binding was analyzed in terms of lifetimes of the transient contacts between 10074-G5 and A β , finding that the interactions were typically shorter than 10 ns, which is a timescale compatible with a diffusion of the small molecule on the fluctuating surface of the disordered peptide. It was also observed that the conformational entropy of A β increased upon interacting with 10074-G5, in a mechanism known as entropic expansion [29], suggesting that exploiting this phenomenon may be a potential therapeutic strategy for disordered proteins. We should note, however, that 10074-G5 is not a specific inhibitor of A β aggregation, since this compound was originally identified using a yeast two-hybrid assay to target the heterodimerisation of c-Myc, a transcription factor involved in cell cycle progression, cellular growth and metabolism, differentiation, and apoptosis, with its partner Max [86], and shown to act by binding and stabilizing the disordered c-Myc monomer [66].

By using NMR spectroscopy, another small molecule, fasudil, has been reported to bind the disordered native state of α -synuclein and inhibit its aggregation [87]. The disordered nature of the complex between fasudil and α -synuclein has been described using molecular dynamics simulations, which revealed that fasudil shuttles between transient networks of side-chains, thus forming short-lived contacts with the protein [65]. More recently, an analysis of these simulations indicated that the binding increases the entropy of α -synuclein [88], consistently with the entropic expansion mechanism of disordered binding [29]. We note that the binding of a small molecule to a disordered protein shifts the populations of its substates [71]. This population shift always underlies an entropic expansion, but it could also leave the entropy unchanged, or reduce it [30].

The disordered N-terminal transactivation domain (NTD) of the androgen receptor (AR), a nuclear receptor involved in most prostate cancers [89], was recently targeted with a series of bisphenol A derivatives, one of which, known as EPI-7386, is currently in phase I clinical trials [90]. The mechanism of binding of two of these derivatives, EPI-002 and EPI-7120, was studied using NMR spectroscopy and molecular dynamics simulations. The two compounds were observed to promote the formation of transient local α -helical structures. Although the bound state remains highly heterogeneous, the modulation of its conformational behavior upon binding is sufficient to prevent its interaction with the transcriptional machinery required for elevated AR transactivation in prostate cancer patients [70].

Taken together, these results indicate that small molecules could be found against the thermodynamic driving force

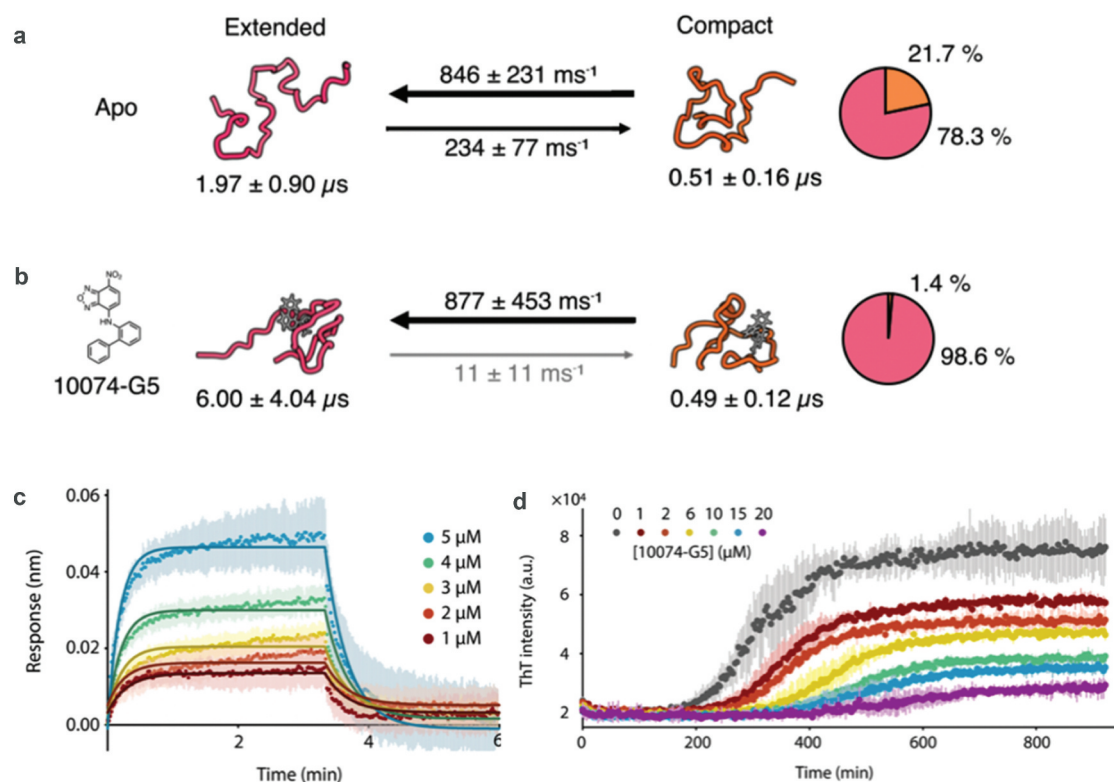


Figure 2. Illustration of a thermodynamic approach. (A) the small molecule 10074-G4 binds A β in its monomeric disordered state [31]. (B) The binding increases the population of this state with respect to more compact, partially structured states on pathway to aggregation. Complex formation takes place through a 'disordered binding mechanism,' since both the peptide and the small molecule remain highly disordered in the bound state [31,32]. Panels a and B are adapted from [31] with permission of American Chemical Society. (C) The dissociation constant can be estimated by surface plasmon resonance (SPR) to be around 6 μM . (D) When present in an in vitro aggregation experiment, 10074-G5 increases in a concentration-dependent manner the amount of monomeric A β at the end of the aggregation reaction. Panels C and D are adapted from [32] with permission of the American Association for the Advancement of Science Publishing Group.

toward protein aggregation. These small molecules are rather effective in solubilizing the native states of proteins, thus reducing their aggregation. At present, however, in the case of disordered proteins, it remains to be established whether it will be possible to obtain in this way small molecules with sufficient affinity and specificity for their targets to be taken forward in drug discovery programs.

3. Kinetic approaches

Kinetic strategies, which are based on the reduction of the rate of aggregation of proteins, require compounds that increase the height of the free energy barrier between the native and amyloid states. A major challenge of this approach is that the one-dimensional free energy barrier shown in Figure 1 is a simplification. More accurately, the macroscopic rate of aggregation is the result of the combination of the rate constants of a complex network of different microscopic processes [33,91,92] (Figure 3A). These microscopic processes include: (i) primary nucleation, which generates disordered oligomeric intermediates from monomeric reagents, (ii) oligomer conversion, which produces ordered oligomers with the characteristic cross- β structure of amyloid fibrils, (iii) elongation, in which amyloid fibrils grow by monomer addition, and (iv) secondary nucleation, where the surfaces of existing amyloid fibrils catalyze the formation of new oligomers [59,95] (Figure 3A).

In this kinetic network, the aggregating proteins are simultaneously the reactants (the monomers), the products (the amyloid fibrils), the intermediates (the oligomers) and the catalysts (the amyloid fibril surfaces). It is thus important to identify the specific microscopic steps that contribute most to the generation of cytotoxic species [96,97]. In some cases, for example, in amyloid light chain amyloidosis, the cytotoxic species are the amyloid fibrils [2,13], while in other cases, for example, for A β in Alzheimer's disease, the most dangerous species are the oligomeric intermediates [1–3,6,23,24]. The non-linear and interconnected nature of the kinetic network underlying A β aggregation makes it difficult to predict the effects of compounds that change the rate constants of specific microscopic steps on the production of oligomers [97] (Figure 3B, C). For example, reducing specifically primary nucleation delays the production of oligomers, but it may not change the total amount of oligomers produced at the end of the reaction. Drugs that inhibit primary nucleation would thus be preventative [34].

It is a major challenge to translate macroscopic measurements on the amounts of amyloid aggregates into estimates of the number of oligomers produced, and hence of whether a drug candidate could be expected to reduce the oligomer-induced toxicity. This is illustrated by the case of compounds that reduce the elongation rate. These compounds delay the formation of amyloid fibrils, but increase the amount of

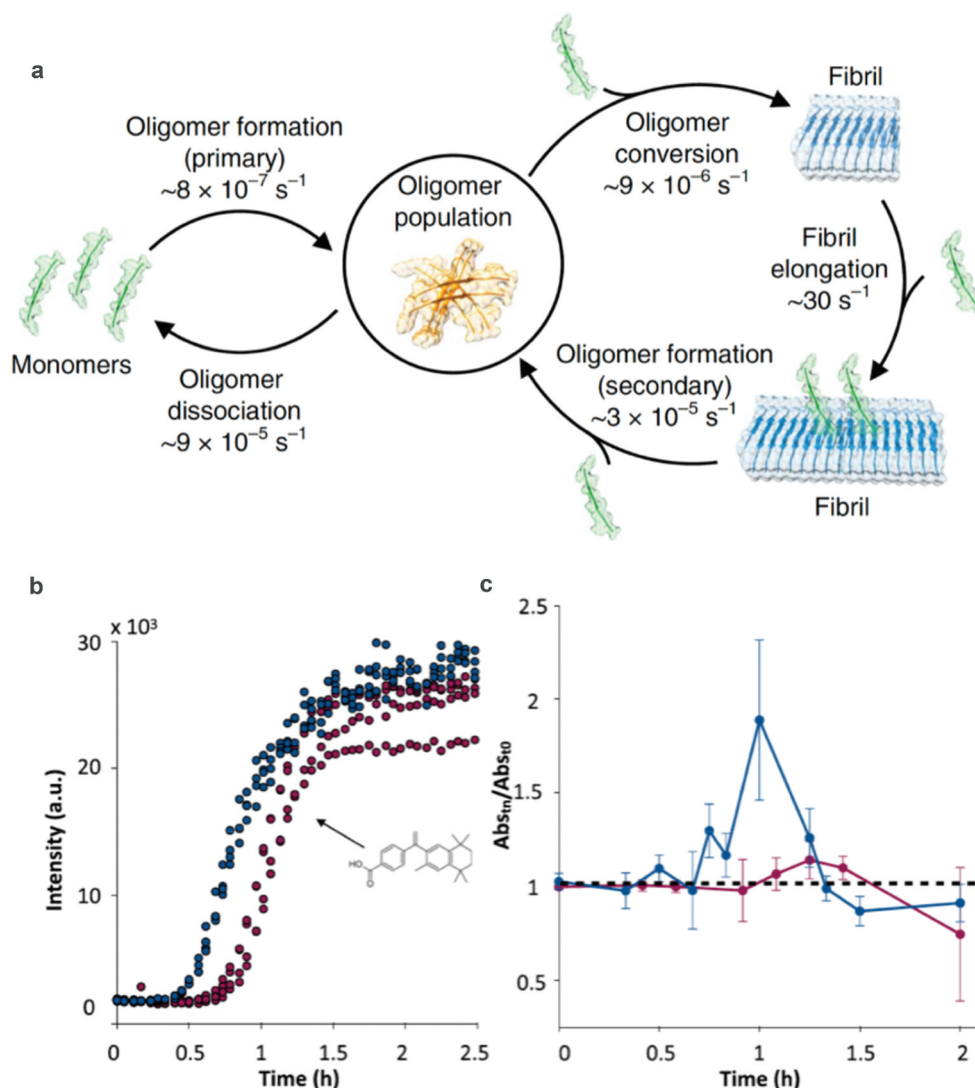


Figure 3. Illustration of a kinetic approach. (A) The free energy barrier in Figure 1 is the result of a complex network of microscopic processes, including primary nucleation, oligomer conversion and dissolution, elongation and secondary nucleation. Adapted from [93] with permission of Springer Nature. (B,C) Small molecules that act on these microscopic processes can delay the formation of amyloid fibrils (B), and reduce the number of oligomers produced during the aggregation reaction (C). Panels B and C are adapted from [94] with permission of the National Academy of Sciences.

oligomers produced during the reaction [97]. This is because the amyloid fibrils act as a sink for monomers. Preventing monomers from binding the fibrils redirects them toward the formation of oligomers. The recent failure in the clinical trials of gantenerumab, an antibody developed to target A β aggregation [98], may at least in part be rationalized in this way [96].

Since in the case of A β and α -synuclein it has been shown that secondary nucleation is the microscopic process that is most effective in producing oligomers [93,99,100], effective compounds could reduce the rate of secondary nucleation [35,36,101]. One way to do this is to use compounds that bind the catalytic sites on the surface of amyloid fibrils. The molecular chaperone Brichos has been shown to act specifically in this way [102]. The antibody aducanumab, the first disease-modifying drug approved by the FDA for Alzheimer's disease [103,104], in addition to its normal effector function, has been shown to have this mechanism of action [96].

Strategies that exploit structure-based drug discovery methods can be applied to the identification and optimization of compounds that bind structured pockets on the surface of amyloid fibrils. If these binding pockets are the catalytic sites for secondary nucleation, the compounds can be effective inhibitors of both amyloid aggregation and oligomer production, as shown recently in the case of α -synuclein aggregation [38]. Machine learning methods are emerging as a promising route for the discovery of potent compounds with this mechanism of action [105,106].

4. Protein aggregation within protein condensates

In the last decade or so, evidence has been accumulating that in addition to the native and amyloid states, proteins can populate a dense, liquid-like state, known as the droplet state, through protein phase separation [81–85,107,108]. The presence of the droplet state opens the possibility of an

alternative pathway to amyloid aggregation, which goes through liquid-like condensates [109–115] (Figure 4A). This process is known as the condensation pathway, while the direct formation of amyloid aggregates from the native state is known as the deposition pathway [116]. The formation of Lewy bodies in Parkinson's disease may take place through either pathway [116]. In ALS, the progressive maturation of liquid-like condensates of FUS and TDP-43 into gels and amyloid aggregates may generate pathological processes either directly through cytotoxic effects of the aggregates or indirectly by preventing the dissolution of the condensates after they have completed their physiological functions [111–115,118].

While many therapeutic opportunities to target the protein phase separation process may be pursued [85,119,120], here we comment on those that concern the inhibition of the amyloid aggregation process within liquid-like condensates. Initial evidence indicates that the type of kinetic

models developed for the deposition pathway may be applicable also to the condensation pathway [117] (Figure 4B). Therefore, the kinetic approach for drug discovery, which was originally developed for the deposition pathway, may conceivably be extended also to the condensation pathway. The thermodynamic approach should also be applicable, since the stabilization of the native state through the binding of small molecules to the native state should reduce the populations of both the condensed and the amyloid states.

5. Conclusions

Although the process of protein misfolding and aggregation remains a challenging target for drug discovery, major advances have been made in recent years, in particular through the validation of the amyloid hypothesis [5,6,104,121,122]. With the first disease-modifying drugs

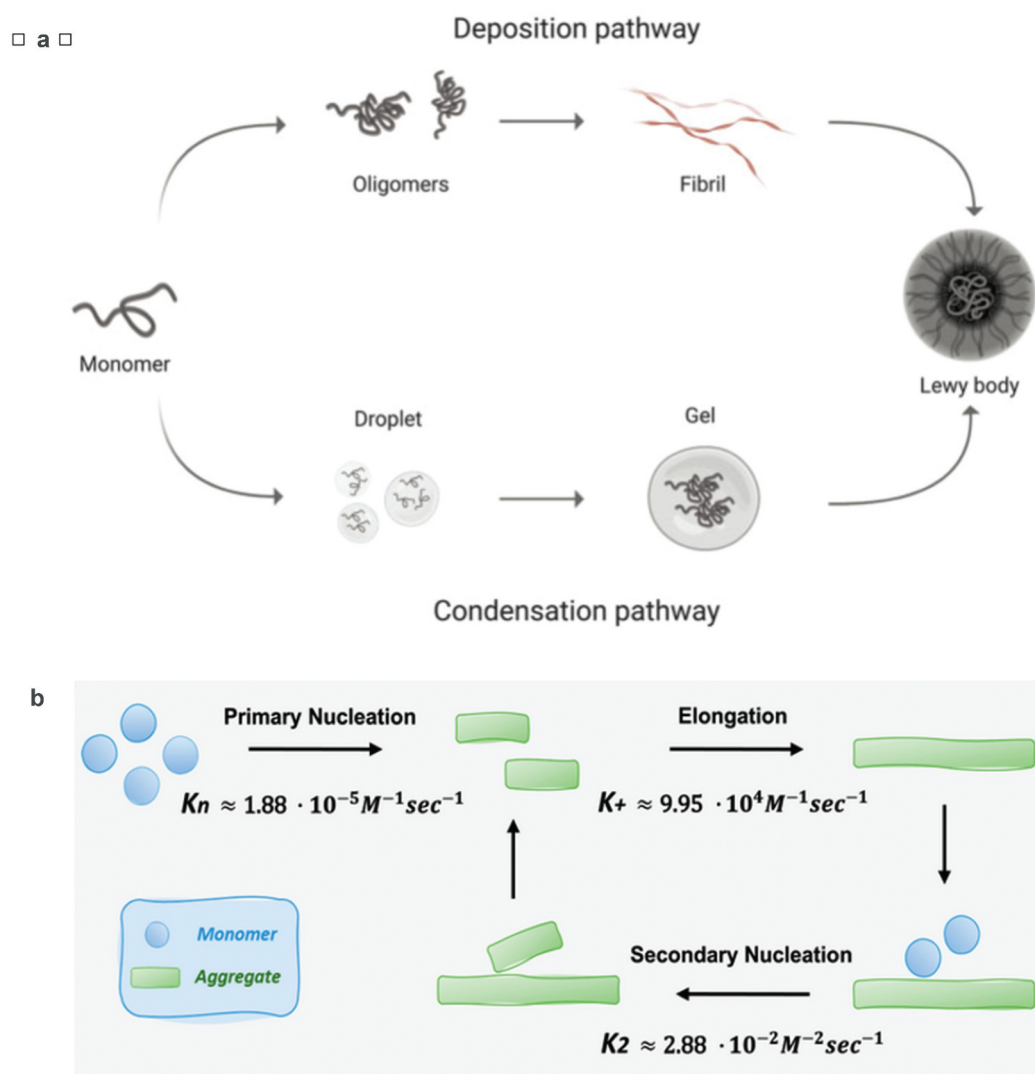


Figure 4. Deposition and condensation pathways in α -synuclein aggregation leading to Lewy body formation. (A) The formation of the amyloid state directly from the native state is known as the deposition pathway, while when it happens through the droplet state is referred to as the condensation pathway. Adapted from [116] with permission of Oxford Academic. (B) In the case of α -synuclein, the kinetic network shown in Figure 3A for the deposition pathway appears to model also the condensation pathway. However, because of the much higher concentration of α -synuclein within condensates, both primary and secondary nucleation are much faster and can be observed in the absence of lipid membranes and at physiological pH along the condensation pathway. Adapted from [117] with permission of the National Academy of Sciences.

having now entered clinical use, the development of more effective compounds with similar targets and mechanisms of action is under way. Both thermodynamic approaches, based on the stabilization of native states [28,53–60], and kinetic approaches, based on the reduction of the rate of aggregation [36,96], are offering powerful routes for establishing successful therapeutic interventions.

6. Expert opinion

There is an urgent need for effective drugs for protein misfolding diseases, which are still largely incurable despite being among the primary causes of disability and death worldwide [123–126]. Drug discovery for these conditions has been extraordinarily challenging, with very few disease-modifying drugs approved to date, and extremely high attrition rates [103,122,127,128]. This situation has been caused, at least in part, by an incomplete understanding of the mechanisms of toxicity in these diseases, and by a lack of effective tools for developing compounds capable of preventing, delaying, or reversing the aggregation process.

On the bright side, the first drugs for protein misfolding diseases are already in clinical use. To date, there are six approved compounds with a mechanism of action based on thermodynamics (Figures 1 and 2) (tafamidis [28,53], migalastat [56], voxelotor [57], ivacaftor [58], elxacaftor [59] and tezacaftor [60]). Two antibodies, aducanumab [103,104] and lecanemab [122], have been approved by the FDA for Alzheimer's disease, which, in addition to their effector functions, are also likely to act through a kinetic mechanism of inhibition of protein aggregation (Figures 1 and 3) [96,121]. These antibodies provide an example of compounds that reduce the proliferation of A β oligomers by inhibiting secondary nucleation [96], and slow down the rate of cognitive decline [122]. The approval of these two antibodies offers support for the amyloid hypothesis [5,6], thus prompting renewed efforts in drug discovery programs based on this concept. The development of small molecules with a similar mechanism of action of aducanumab and lecanemab may result in fewer adverse reactions, and, in the case of intracellular aggregates, such as tau tangles and α -synuclein Lewy bodies, in better target engagement.

To extend these advances to other protein misfolding diseases, it will be important to develop more accurate methods for the identification and the quantitative measurements of the most toxic species produced when proteins misfold and aggregate. This is a challenging task, since the aggregation process involves many interconverting species, which are difficult to separate and characterize individually within a complex kinetic network [93,95,96,129]. This task is intertwined with the development of pre-clinical toxicity assays that recapitulate the pathological mechanisms relevant in a disease. Neuronal cell models derived from induced pluripotent stem cells hold great promise in this direction [130–132], as also indicated by the recent revision of the FDA guidelines concerning the use of animal models.

Diagnostic methods, in particular, based on biofluids should be developed in parallel to drug discovery [133]. These methods are essential to assess the clinical efficacy of treatments, particularly those aimed at reducing the amounts of oligomers

produced during an aggregation reaction, which are highly elusive species [129]. Accurate oligomer detection and quantification methods are also going to be necessary in pre-clinical target engagement studies [94,96,99,129]. Such methods are also required for the systematic optimization of the potency of compounds during the early stages of drug discovery, where structure–activity relationships should be developed [36,93].

Moving forward, the introduction of machine learning methods is going to facilitate the drug discovery process at preclinical stages, as is already happening for other diseases, by reducing through rapid computational screening the time required for hit identification and lead optimization, as well as for accurate predictions of pharmacokinetics, pharmacodynamics, and toxicity [134,135].

An area where exciting progress may be expected is the targeting of disordered proteins by small molecules. Lessons learned from the thermodynamic approaches described here could have a major impact in drug discovery programs for many major human conditions where disordered proteins play central roles related to the dysregulation of their native states [29,67–69].

Another direction where major advances are likely to emerge in the coming decade is the extension of the type of drug discovery described in this review to protein condensation diseases, where a dysregulation of the balance between the native, droplet, and amyloid state appears to underlie many pathological processes [85,119,120].

Funding

M Vendruscolo is supported by UK Research and Innovation through grants 10061100 and 10059436 and the Wellcome Trust through grant 203249/Z/16/Z.

Declaration of interest

M Vendruscolo is a founder of Wren Therapeutics (now WaveBreak Therapeutics). The author has no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

Reviewer disclosures

Peer reviewers on this manuscript have no relevant financial or other relationships to disclose.

ORCID

Michele Vendruscolo  <http://orcid.org/0000-0002-3616-1610>

References

Papers of special note have been highlighted as: (•) of interest (••) of considerable interest

- Knowles TP, Vendruscolo M, Dobson CM. The amyloid state and its association with protein misfolding diseases. *Nat Rev Mol Cell Biol.* 2014;15(6):384–396.
- Merlini G, Bellotti V. Molecular mechanisms of amyloidosis. *N Engl J Med.* 2003;349(6):583–596.
- Buxbaum JN, Dispenzieri A, Eisenberg DS, et al. Amyloid nomenclature 2022: update, novel proteins, and recommendations by the International Society of Amyloidosis (ISA) nomenclature

- committee. Amyloid. 2022;29(4):213–219. DOI:10.1080/13506129.2022.2147636
4. Dobson CM. Protein folding and misfolding. *Nature*. 2003;426(6968):884–890.
5. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med*. 2016;8(6):595–608.
6. Hampel H, Hardy J, Blennow K, et al. The amyloid- β pathway in Alzheimer's disease. *Mol Psychiatry*. 2021;26(10):5481–5503. DOI:10.1038/s41380-021-01249-0
7. Grundke-Iqbal I, Iqbal K, Tung Y-C, et al. Abnormal phosphorylation of the microtubule-associated protein tau (τ) in Alzheimer cytoskeletal pathology. *Proc Natl Acad Sci USA*. 1986;83(13):4913–4917. DOI:10.1073/pnas.83.13.4913
8. Lee VM, Goedert M, Trojanowski JQ. Neurodegenerative tauopathies. *Annu Rev Neurosci*. 2001;24:1121.
9. Spillantini MG, Schmidt ML, Lee V-Y, et al. α -synuclein in Lewy bodies. *Nature*. 1997;388(6645):839–840. DOI:10.1038/42166
10. Baba M, Nakajo S, Tu P-H, et al. Aggregation of α -synuclein in Lewy bodies of sporadic Parkinson's disease and dementia with Lewy bodies. *Am J Pathol*. 1998;152(4):879.
11. Westermarck P, Wernstedt C, Wilander E, et al. A novel peptide in the calcitonin gene related peptide family as an amyloid fibril protein in the endocrine pancreas. *Biochem Biophys Res Commun*. 1986;140(3):827–831. DOI:10.1016/0006-291X(86)90708-4
12. Cooper G, Willis A, Clark A, et al. Purification and characterization of a peptide from amyloid-rich pancreases of type 2 diabetic patients. *Proc Natl Acad Sci USA*. 1987;84(23):8628–8632. DOI:10.1073/pnas.84.23.8628
13. Falk RH, Alexander KM, Liao R, et al. AL (light-chain) cardiac amyloidosis: a review of diagnosis and therapy. *J Am Coll Cardiol*. 2016;68(12):1323–1341. DOI:10.1016/j.jacc.2016.06.053
14. Gejyo F, Yamada T, Odani S, et al. A new form of amyloid protein associated with chronic hemodialysis was identified as β 2-microglobulin. *Biochem Biophys Res Commun*. 1985;129(3):701–706. DOI:10.1016/0006-291X(85)91948-5
15. Ando Y, Nakamura M, Araki S. Transthyretin-related familial amyloidotic polyneuropathy. *Arch Neurol*. 2005;62(7):1057–1062.
16. Chiti F, Kelly JW. Small molecule protein binding to correct cellular folding or stabilize the native state against misfolding and aggregation. *Curr Opin Struct Biol*. 2022;72:267–278.
17. Eisenberg D, Jucker M. The amyloid state of proteins in human diseases. *Cell*. 2012;148(6):1188–1203.
18. Knowles TP, Fitzpatrick AW, Meehan S, et al. Role of intermolecular forces in defining material properties of protein nanofibrils. *Science*. 2007;318(5858):1900–1903. DOI:10.1126/science.1150057
19. Yang Y, Shi Y, Schweighauser M, et al. Structures of α -synuclein filaments from human brains with Lewy pathology. *Nature*. 2022;610(7933):791–795. DOI:10.1038/s41586-022-05319-3
20. Fitzpatrick AW, Falcon B, He S, et al. Cryo-EM structures of tau filaments from Alzheimer's disease. *Nature*. 2017;547(7662):185–190. DOI:10.1038/nature23002
21. Yang Y, Arseni D, Zhang W, et al. Cryo-EM structures of amyloid- β 42 filaments from human brains. *Science*. 2022;375(6577):167–172. DOI:10.1126/science.abm7285
22. Shi Y, Zhang W, Yang Y, et al. Structure-based classification of tauopathies. *Nature*. 2021;598(7880):359–363. DOI:10.1038/s41586-021-03911-7
23. Haass C, Selkoe DJ. Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid β -peptide. *Nat Rev Mol Cell Biol*. 2007;8(2):101–112.
24. Benilova I, Karran E, De Strooper B. The toxic A β oligomer and Alzheimer's disease: an emperor in need of clothes. *Nat Neurosci*. 2012;15(3):349–357.
25. Ciryam P, Tartaglia GG, Morimoto RI, et al. Widespread aggregation and neurodegenerative diseases are associated with supersaturated proteins. *Cell Rep*. 2013;5(3):781–790. DOI:10.1016/j.celrep.2013.09.043
26. Vecchi G, Sormanni P, Mannini B, et al. Proteome-wide observation of the phenomenon of life on the edge of solubility. *Proc Natl Acad Sci USA*. 2020;117(2):1015–1020. DOI:10.1073/pnas.1910444117
27. Tartaglia GG, Pechmann S, Dobson CM, et al. Life on the edge: a link between gene expression levels and aggregation rates of human proteins. *Trends Biochem Sci*. 2007;32(5):204–206. DOI:10.1016/j.tibs.2007.03.005
28. Miroy GJ, Lai Z, Lashuel HA, et al. Inhibiting transthyretin amyloid fibril formation via protein stabilization. *Proc Natl Acad Sci USA*. 1996;93(26):15051–15056. DOI:10.1073/pnas.93.26.15051
- **This seminal article describes the thermodynamic strategy of stabilizing the native state of a protein to prevent its aggregation.**
29. Heller GT, Sormanni P, Vendruscolo M. Targeting disordered proteins with small molecules using entropy. *Trends Biochem Sci*. 2015;40(9):491–496.
30. Heller GT, Bonomi M, Vendruscolo M. Structural ensemble modulation upon small-molecule binding to disordered proteins. *J Mol Biol*. 2018;430(16):2288–2292.
31. Lohr T, Kohlhoff K, Heller GT, et al. A small molecule stabilizes the disordered native state of the Alzheimer's A β peptide. *ACS Chem Neurosci*. 2022;13(12):1738–1745. DOI:10.1021/acscchemneuro.2c00116
32. Heller GT, Aprile FA, Michaels TC, et al. Small-molecule sequestration of amyloid- β as a drug discovery strategy for Alzheimer's disease. *Sci Adv*. 2020;6(45):eabb5924. DOI:10.1126/sciadv.abb5924
- **This article reports an initial example of the thermodynamic strategy for a disordered protein.**
33. Arosio P, Vendruscolo M, Dobson CM, et al. Chemical kinetics for drug discovery to combat protein aggregation diseases. *Trends Pharmacol Sci*. 2014;35(3):127–135. DOI:10.1016/j.tips.2013.12.005
34. Habchi J, Arosio P, Perni M, et al. An anticancer drug suppresses the primary nucleation reaction that initiates the production of the toxic A β 42 aggregates linked with Alzheimer's disease. *Sci Adv*. 2016;2(2):e1501244. DOI:10.1126/sciadv.1501244
- **This article illustrates the use of the kinetic approach to inhibit the rate of primary nucleation.**
35. Habchi J, Chia S, Limbicker R, et al. Systematic development of small molecules to inhibit specific microscopic steps of A β 42 aggregation in Alzheimer's disease. *Proc Natl Acad Sci USA*. 2017;114(2):E200–E208. DOI:10.1073/pnas.1615613114
36. Chia S, Habchi J, Michaels TC, et al. SAR by kinetics for drug discovery in protein misfolding diseases. *Proc Natl Acad Sci USA*. 2018;115(41):10245–10250. DOI:10.1073/pnas.1807884115
- **This article describes how the use the kinetic approach to reduce the rate of production of misfolded protein oligomers as an optimization parameter for drug discovery.**
37. Michaels TC, Šarić A, Meisl G, et al. Thermodynamic and kinetic design principles for amyloid-aggregation inhibitors. *Proc Natl Acad Sci USA*. 2020;117(39):24251–24257. DOI:10.1073/pnas.2006684117
38. Chia S, Brotzakis ZF, Horne RI, et al. Structure-based discovery of small molecule inhibitors of the autocatalytic proliferation of α -synuclein aggregates. *Mol Pharm*. 2023;20(1):183–193. DOI:10.1021/acs.molpharmaceut.2c00548
39. Baldwin AJ, Knowles TP, Tartaglia GG, et al. Metastability of native proteins and the phenomenon of amyloid formation. *J Am Chem Soc*. 2011;133(36):14160–14163. DOI:10.1021/ja2017703
40. Kundra R, Ciryam P, Morimoto RI, et al. Protein homeostasis of a metastable subproteome associated with Alzheimer's disease. *Proc Natl Acad Sci USA*. 2017;114(28):E5703–E5711. DOI:10.1073/pnas.1618417114
41. Balch WE, Morimoto RI, Dillin A, et al. Adapting proteostasis for disease intervention. *Science*. 2008;319(5865):916–919. DOI:10.1126/science.1141448
42. Hipp MS, Kasturi P, Hartl FU. The proteostasis network and its decline in ageing. *Nat Rev Mol Cell Biol*. 2019;20(7):421–435.
43. Kelmer Sacramento E, Kirkpatrick JM, Mazzetto M, et al. Reduced proteasome activity in the aging brain results in ribosome stoichiometry loss and aggregation. *Mol Sys Biol*. 2020;16(6):e9596. DOI:10.15252/msb.20209596
44. Huang C, Wagner-Valladolid S, Stephens AD, et al. Intrinsically aggregation-prone proteins form amyloid-like aggregates and

- contribute to tissue aging in *Caenorhabditis elegans*. *Elife*. 2019;8:e43059.
45. Chen YR, Harel I, Singh PP, et al. Tissue-specific landscape of protein aggregation and quality control in an aging vertebrate. *bioRxiv*. 2022. DOI:10.1101/2022.02.26.482120
 46. Harel I, Chen YR, Ziv I, et al. Identification of protein aggregates in the aging vertebrate brain with prion-like and phase separation properties. *bioRxiv*. 2022. DOI:10.1101/2022.02.26.482115
 47. David DC, Ollikainen N, Trinidad JC, et al. Widespread protein aggregation as an inherent part of aging in *C. elegans*. *PLoS Biol*. 2010;8(8):e1000450. DOI:10.1371/journal.pbio.1000450
 48. Walther DM, Kasturi P, Zheng M, et al. Widespread proteome remodeling and aggregation in aging *C. elegans*. *Cell*. 2015;161(4):919–932. DOI:10.1016/j.cell.2015.03.032
 49. Sui X, Pires DE, Ormsby AR, et al. Widespread remodeling of proteome solubility in response to different protein homeostasis stresses. *Proc Natl Acad Sci USA*. 2020;117(5):2422–2431. DOI:10.1073/pnas.1912897117
 50. Copeland RA, Pompliano DL, Meek TD. Drug–target residence time and its implications for lead optimization. *Nat Rev Drug Discov*. 2006;5(9):730–739.
 51. Pan AC, Borhani DW, Dror RO, et al. Molecular determinants of drug–receptor binding kinetics. *Drug Discov Today*. 2013;18(13–14):667–673. DOI:10.1016/j.drudis.2013.02.007
 52. Liu W, Jiang J, Lin Y, et al. Insight into thermodynamic and kinetic profiles in small-molecule optimization. *J Med Chem*. 2022;65(16):10809–10847. DOI:10.1021/acs.jmedchem.2c00682
 53. Bulawa CE, Connelly S, DeVit M, et al. Tafamidis, a potent and selective transthyretin kinetic stabilizer that inhibits the amyloid cascade. *Proc Natl Acad Sci USA*. 2012;109(24):9629–9634. DOI:10.1073/pnas.1121005109
 54. Hammarstrom P, Wiseman RL, Powers ET, et al. Prevention of transthyretin amyloid disease by changing protein misfolding energetics. *Science*. 2003;299(5607):713–716. DOI:10.1126/science.1079589
 55. Ringe D, Petsko GA. Q&A: what are pharmacological chaperones and why are they interesting? *J Biol*. 2009;8(9):1–4.
 56. Fan J-Q, Ishii S, Asano N, et al. Accelerated transport and maturation of lysosomal α -galactosidase in Fabry lymphoblasts by an enzyme inhibitor. *Nat Med*. 1999;5(1):112–115. DOI:10.1038/4801
 57. Vichinsky E, Hoppe CC, Ataga KI, et al. A phase 3 randomized trial of voxelotor in sickle cell disease. *N Engl J Med*. 2019;381(6):509–519. DOI:10.1056/NEJMoa1903212
 58. Ramsey BW, Davies J, McElvaney NG, et al. A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N Engl J Med*. 2011;365(18):1663–1672. DOI:10.1056/NEJMoa1105185
 59. Middleton PG, Mall MA, Dřevínek P, et al. Elexacaftor–tezacaftor–ivacaftor for cystic fibrosis with a single Phe508del allele. *N Engl J Med*. 2019;381(19):1809–1819. DOI:10.1056/NEJMoa1908639
 60. Taylor-Cousar JL, Munck A, McKone EF, et al. Tezacaftor–ivacaftor in patients with cystic fibrosis homozygous for Phe508del. *N Engl J Med*. 2017;377(21):2013–2023. DOI:10.1056/NEJMoa1709846
 61. Makley LN, McMenimen KA, DeVree BT, et al. Pharmacological chaperone for α -crystallin partially restores transparency in cataract models. *Science*. 2015;350(6261):674–677. DOI:10.1126/science.aac9145
 62. Molnar KS, Donyak BM, Su B, et al. Mechanism of action of VP1-001 in cryAB (R120G)-associated and age-related cataracts. *Invest Ophthalmol Vis Sci*. 2019;60(10):3320–3331. DOI:10.1167/iov.18-25647
 63. Zhao L, Chen X-J, Zhu J, et al. Lanosterol reverses protein aggregation in cataracts. *Nature*. 2015;523(7562):607–611. DOI:10.1038/nature14650
 64. Stephenson Clarke JR, Douglas LR, Duriez PJ, et al. Discovery of nanomolar-affinity pharmacological chaperones stabilizing the oncogenic p53 mutant Y220C. *ACS Pharmacol Transl Sci*. 2022;5(11):1169–1180. DOI:10.1021/acsp.2c00164
 65. Robustelli P, Ibanez-de-Opakua A, Campbell-Bezatz C, et al. Molecular basis of small-molecule binding to α -synuclein. *J Am Chem Soc*. 2022;144(6):2501–2510. DOI:10.1021/jacs.1c07591
 66. Follis AV, Hammoudeh DI, Wang H, et al. Structural rationale for the coupled binding and unfolding of the c-Myc oncoprotein by small molecules. *Chem Biol*. 2008;15(11):1149–1155. DOI:10.1016/j.chembiol.2008.09.011
 67. Uversky VN. Intrinsically disordered proteins and novel strategies for drug discovery. *Expert Opin Drug Discov*. 2012;7(6):475–488.
 68. Biesaga M, Frigolé-Vivas M, Salvatella X. Intrinsically disordered proteins and biomolecular condensates as drug targets. *Curr Opin Chem Biol*. 2021;62:90–100.
 69. Ruan H, Sun Q, Zhang W, et al. Targeting intrinsically disordered proteins at the edge of chaos. *Drug Discov Today*. 2019;24(1):217–227. DOI:10.1016/j.drudis.2018.09.017
 70. Zhu J, Salvatella X, Robustelli P. Small molecules targeting the disordered transactivation domain of the androgen receptor induce the formation of collapsed helical states. *Nat Comm*. 2022;13(1):1–15.
 71. Ban D, Iconaru LI, Ramanathan A, et al. A small molecule causes a population shift in the conformational landscape of an intrinsically disordered protein. *J Am Chem Soc*. 2017;139(39):13692–13700. DOI:10.1021/jacs.7b01380
 72. Ruan H, Yu C, Niu X, et al. Computational strategy for intrinsically disordered protein ligand design leads to the discovery of p53 transactivation domain I binding compounds that activate the p53 pathway. *Chem Sci*. 2021;12(8):3004–3016. DOI:10.1039/D0SC04670A
 73. Zhu M, De Simone A, Schenk D, et al. Identification of small-molecule binding pockets in the soluble monomeric form of the A β 42 peptide. *J Chem Phys*. 2013;139(3):07B609_1. DOI:10.1063/1.4811831
 74. Toth G, Gardai SJ, Zago W, et al. Targeting the intrinsically disordered structural ensemble of α -synuclein by small molecules as a potential therapeutic strategy for Parkinson's disease. *PLoS One*. 2014;9(2):e87133. DOI:10.1371/journal.pone.0087133
 75. Ball SR, Adamson JS, Sullivan MA, et al. Perphenazine–macrocyclic conjugates rapidly sequester the A β 42 monomer and prevent formation of toxic oligomers and amyloid. *ACS Chem Neurosci*. 2022;14(1):87–98. DOI:10.1021/acscchemneuro.2c00498
 76. Leguizamón Herrera VL, Buell AK, Willbold D, et al. Interaction of therapeutic d-peptides with A β 42 monomers, thermodynamics, and binding analysis. *ACS Chem Neurosci*. 2022;13(11):1638–1650. DOI:10.1021/acscchemneuro.2c00102
 77. Gallego-Villarejo L, Wallin C, Król S, et al. Big dynorphin is a neuroprotector scaffold against amyloid β -peptide aggregation and cell toxicity. *Comput Struct Biotechnol J*. 2022;20:5672–5679.
 78. Borgia A, Borgia MB, Bugge K, et al. Extreme disorder in an ultrahigh-affinity protein complex. *Nature*. 2018;555(7694):61–66. DOI:10.1038/nature25762
 79. Sottini A, Borgia A, Borgia MB, et al. Polyelectrolyte interactions enable rapid association and dissociation in high-affinity disordered protein complexes. *Nat Comm*. 2020;11(1):1–14. DOI:10.1038/s41467-020-18859-x
 80. Schuler B, Borgia A, Borgia MB, et al. Binding without folding—the biomolecular function of disordered polyelectrolyte complexes. *Curr Opin Struct Biol*. 2020;60:66–76.
 81. Banani SF, Lee HO, Hyman AA, et al. Biomolecular condensates: organizers of cellular biochemistry. *Nat Rev Mol Cell Biol*. 2017;18(5):285–298. DOI:10.1038/nrm.2017.7
 82. Shin Y, Brangwynne CP. Liquid phase condensation in cell physiology and disease. *Science*. 2017;357(6357):eaaf4382.
 83. Alberti S, Hyman AA. Biomolecular condensates at the nexus of cellular stress, protein aggregation disease and ageing. *Nat Rev Mol Cell Biol*. 2021;22(3):196–213.
 84. Fuxreiter M, Vendruscolo M. Generic nature of the condensed states of proteins. *Nat Cell Biol*. 2021;23(6):587–594.
 85. Vendruscolo M, Fuxreiter M. Protein condensation diseases: therapeutic opportunities. *Nat Comm*. 2022;13(1):1–11.
 86. Yin X, Giap C, Lazo JS, et al. Low molecular weight inhibitors of Myc–max interaction and function. *Oncogene*. 2003;22(40):6151–6159. DOI:10.1038/sj.onc.1206641

87. Tatenhorst L, Eckermann K, Dambeck V, et al. Fasudil attenuates aggregation of α -synuclein in models of Parkinson's disease. *Acta Neuropathol Commun.* 2016;4(1):1–17. DOI:10.1186/s40478-016-0310-y
88. Menon S, Mondal J. Conformational ensemble of monomeric α -synuclein in aqueous and crowded environments as revealed by markov state model. *bioRxiv.* 2022. DOI:10.1101/2022.02.20.481191
89. Sadar MD. Discovery of drugs that directly target the intrinsically disordered region of the androgen receptor. *Expert Opin Drug Discov.* 2020;15(5):551–560.
90. Hong NH, Le Moigne R, Pearson P, et al. The preclinical characterization of the N-terminal domain androgen receptor inhibitor, EPI-7386, for the treatment of prostate cancer. *Eur J Cancer.* 2020;138:551.
91. Cohen SI, Vendruscolo M, Dobson CM, et al. From macroscopic measurements to microscopic mechanisms of protein aggregation. *J Mol Biol.* 2012;421(2–3):160–171. DOI:10.1016/j.jmb.2012.02.031
92. Meisl G, Kirkegaard JB, Arosio P, et al. Molecular mechanisms of protein aggregation from global fitting of kinetic models. *Nat Protoc.* 2016;11(2):252–272. DOI:10.1038/nprot.2016.010
93. Michaels TC, Šarić A, Curk S, et al. Dynamics of oligomer populations formed during the aggregation of Alzheimer's $\text{A}\beta$ 42 peptide. *Nat Chem.* 2020;12(5):445–451. DOI:10.1038/s41557-020-0452-1
94. Aprile FA, Sormanni P, Podpolny M, et al. Rational design of a conformation-specific antibody for the quantification of $\text{A}\beta$ oligomers. *Proc Natl Acad Sci USA.* 2020;117(24):13509–13518. DOI:10.1073/pnas.1919464117
95. Knowles TP, Waudby CA, Devlin GL, et al. An analytical solution to the kinetics of breakable filament assembly. *Science.* 2009;326(5959):1533–1537. DOI:10.1126/science.1178250
96. Linse S, Scheidt T, Bernfur K, et al. Kinetic fingerprints differentiate the mechanisms of action of anti- $\text{A}\beta$ antibodies. *Nat Struct Mol Biol.* 2020;27(12):1125–1133. DOI:10.1038/s41594-020-0505-6
- **This article describes the mechanism of action of clinical stage antibodies for Alzheimer's disease in terms of their kinetic mechanism.**
97. Michaels TC, Dear AJ, Cohen SI, et al. Kinetic profiling of therapeutic strategies for inhibiting the formation of amyloid oligomers. *J Chem Phys.* 2022;156(16):164904. DOI:10.1063/5.0077609
98. Ostrowitzki S, Lasser RA, Dorflinger E, et al. A phase III randomized trial of gantenerumab in prodromal Alzheimer's disease. *Alzheimer's Res Ther.* 2017;9(1):1–15. DOI:10.1186/s13195-017-0318-y
99. Cohen SI, Linse S, Luheshi LM, et al. Proliferation of amyloid- β 42 aggregates occurs through a secondary nucleation mechanism. *Proc Natl Acad Sci USA.* 2013;110(24):9758–9763. DOI:10.1073/pnas.1218402110
100. Buell AK, Galvagnion C, Gaspar R, et al. Solution conditions determine the relative importance of nucleation and growth processes in α -synuclein aggregation. *Proc Natl Acad Sci USA.* 2014;111(21):7671–7676. DOI:10.1073/pnas.1315346111
101. Staats R, Michaels TC, Flagmeier P, et al. Screening of small molecules using the inhibition of oligomer formation in α -synuclein aggregation as a selection parameter. *Comm Chem.* 2020;3(1):1–9. DOI:10.1038/s42004-020-00412-y
102. Cohen SI, Arosio P, Presto J, et al. A molecular chaperone breaks the catalytic cycle that generates toxic $\text{A}\beta$ oligomers. *Nat Struct Mol Biol.* 2015;22(3):207–213. DOI:10.1038/nsmb.2971
103. Sevigny J, Chiao P, Bussi re T, et al. The antibody aducanumab reduces $\text{A}\beta$ plaques in Alzheimer's disease. *Nature.* 2016;537(7618):50–56. DOI:10.1038/nature19323
- **This article reports the development of aducanumab, which then became the first disease-modifying drug for Alzheimer's disease approved by the FDA.**
104. Budd Haeberlein S, Aisen P, Barkhof F, et al. Two randomized phase 3 studies of aducanumab in early Alzheimer's disease. *J Prev Alzheimers Dis.* 2022;9(2):197–210. DOI:10.14283/jpad.2022.30
105. Horne R, Possenti A, Chia S, et al. A machine learning approach to identify small molecule inhibitors of secondary nucleation in α -synuclein aggregation. *bioRxiv.* 2021. DOI:10.1101/2021.11.10.468009
106. Horne RI, Murtada MH, Huo D, et al. Exploration and exploitation approaches based on generative machine learning to identify potent small molecule inhibitors of α -synuclein secondary nucleation. *J Chem Theory Comput.* 2023. DOI:10.1021/acs.jctc.2c01303
107. Hardenberg M, Horvath A, Ambrus V, et al. Widespread occurrence of the droplet state of proteins in the human proteome. *Proc Natl Acad Sci USA.* 2020;117(52):33254–33262. DOI:10.1073/pnas.2007670117
108. Brangwynne CP, Eckmann CR, Courson DS, et al. Germline P granules are liquid droplets that localize by controlled dissolution/condensation. *Science.* 2009;324(5935):1729–1732. DOI:10.1126/science.1172046
109. Stender EG, Ray S, Norrild RK, et al. Capillary flow experiments for thermodynamic and kinetic characterization of protein liquid-liquid phase separation. *Nat Comm.* 2021;12(1):1–18. DOI:10.1038/s41467-021-27433-y
110. Ray S, Singh N, Kumar R, et al. α -Synuclein aggregation nucleates through liquid-liquid phase separation. *Nat Chem.* 2020;12(8):705–716. DOI:10.1038/s41557-020-0465-9
111. Han TW, Kato M, Xie S, et al. Cell-free formation of RNA granules: bound RNAs identify features and components of cellular assemblies. *Cell.* 2012;149(4):768–779. DOI:10.1016/j.cell.2012.04.016
112. Patel A, Lee HO, Jawerth L, et al. A liquid-to-solid phase transition of the ALS protein FUS accelerated by disease mutation. *Cell.* 2015;162(5):1066–1077. DOI:10.1016/j.cell.2015.07.047
113. Qamar S, Wang G, Randle SJ, et al. FUS phase separation is modulated by a molecular chaperone and methylation of arginine cation- π interactions. *Cell.* 2018;173(3):720–734. e15. DOI:10.1016/j.cell.2018.03.056
114. Neumann M, Sampathu DM, Kwong LK, et al. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science.* 2006;314(5796):130–133. DOI:10.1126/science.1134108
115. Braak H, Bretschneider J, Ludolph AC, et al. Amyotrophic lateral sclerosis—a model of corticofugal axonal spread. *Nat Rev Neurol.* 2013;9(12):708–714. DOI:10.1038/nrneurol.2013.221
116. Hardenberg MC, Sinnige T, Casford S, et al. Observation of an α -synuclein liquid droplet state and its maturation into Lewy body-like assemblies. *J Mol Cell Biol.* 2021;13(4):282–294. DOI:10.1093/jmcb/mjaa075
117. Dada ST, Hardenberg MC, Mrugalla LK, et al. Spontaneous nucleation and fast aggregate-dependent proliferation of α -synuclein aggregates within liquid condensates at physiological pH. *Proc Natl Acad Sci USA.* 2023;120:e2208792120.
118. Bolognesi B, Faure AJ, Seuma M, et al. The mutational landscape of a prion-like domain. *Nat Comm.* 2019;10(1):1–12. DOI:10.1038/s41467-019-12101-z
119. Mitrea DM, Mittasch M, Gomes BF, et al. Modulating biomolecular condensates: a novel approach to drug discovery. *Nat Rev Drug Discov.* 2022;21(11):841–862. DOI:10.1038/s41573-022-00505-4
120. Berkeley RF, Debelouchina GT. Chemical tools for study and modulation of biomolecular phase transitions. *Chem Sci.* 2022;13(48):14226–14245.
121. S derberg L, Johannesson M, Nygren P, et al. Lecanemab, aducanumab, and gantenerumab—binding profiles to different forms of amyloid-beta might explain efficacy and side effects in clinical trials for Alzheimer's disease. *Neurotherapeutics.* 2022;20(1):195–206. DOI:10.1007/s13311-022-01308-6
122. van Dyck CH, Swanson CJ, Aisen P, et al. Lecanemab in early Alzheimer's disease. *N Engl J Med.* 2022;388(1):9–21. DOI:10.1056/NEJMoa2212948
123. Gustavsson A, Norton N, Fast T, et al. Global estimates on the number of persons across the Alzheimer's disease continuum. *Alzheimer's Dement.* 2022;19(2):658–670. DOI:10.1002/alz.12694

124. Qiu C, Kivipelto M, Von Strauss E. Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. *Dialogues Clin Neurosci*. 2022;11(2): 111–128 .
125. Balestrino R, Schapira A. Parkinson disease. *Eur J Neurol*. 2020;27(1):27–42.
126. Livingston G, Huntley J, Sommerlad A, et al. Dementia prevention, intervention, and care: 2020 report of the Lancet Commission. *Lancet*. 2020;396(10248):413–446. DOI:10.1016/S0140-6736(20)30367-6
127. Cummings J, Lee G, Nahed P, et al. Alzheimer's disease drug development pipeline: 2022. *Alzheimer's Dement*. 2022;8(1): e12295. DOI:10.1002/trc2.12295
128. Cummings JL, Goldman DP, Simmons-Stern NR, et al. The costs of developing treatments for Alzheimer's disease: a retrospective exploration. *Alzheimer's Dement*. 2022;18(3):469–477. DOI:10.1002/alz.12450
129. Kulenkampff K, Wolf Perez A-M, Sormanni P, et al. Quantifying misfolded protein oligomers as drug targets and biomarkers in Alzheimer and Parkinson diseases. *Nat Rev Chem*. 2021;5(4):277–294. DOI:10.1038/s41570-021-00254-9
130. Park J, Wetzel I, Marriott I, et al. A 3D human triculture system modeling neurodegeneration and neuroinflammation in Alzheimer's disease. *Nat Neurosci*. 2018;21(7):941–951. DOI:10.1038/s41593-018-0175-4
131. Lancaster MA, Knoblich JA. Generation of cerebral organoids from human pluripotent stem cells. *Nat Protoc*. 2014;9(10):2329–2340.
132. Paşca AM, Sloan SA, Clarke LE, et al. Functional cortical neurons and astrocytes from human pluripotent stem cells in 3D culture. *Nat Methods*. 2015;12(7):671–678. DOI:10.1038/nmeth.3415
133. Teunissen CE, Verberk IM, Thijssen EH, et al. Blood-based biomarkers for Alzheimer's disease: towards clinical implementation. *Lancet Neurol*. 2022;21(1):66–77. DOI:10.1016/S1474-4422(21)00361-6
134. Jiménez-Luna J, Grisoni F, Weskamp N, et al. Artificial intelligence in drug discovery: recent advances and future perspectives. *Expert Opin Drug Discov*. 2021;16(9):949–959. DOI:10.1080/17460441.2021.1909567
135. Cheng F, Cummings J. Artificial intelligence in Alzheimer's drug discovery. *Alzheimer's Disease Drug Development*. 2022;6:62–72.